Bioorganic & Medicinal Chemistry 14 (2006) 6165-6173

Bioorganic & Medicinal Chemistry

A flexible approach for the preparation of substituted benzazepines: Application to the synthesis of tolvaptan[☆]

Alejandro Cordero-Vargas, Béatrice Quiclet-Sire and Samir Z. Zard*

Laboratoire de Synthèse Organique associé au CNRS, Ecole Polytechnique, 91128 Palaiseau, France

Received 14 February 2006; revised 21 May 2006; accepted 31 May 2006 Available online 16 June 2006

Dedicated with respect to Professor Bernd Giese, recipient of last year's Tetrahedron Prize.

Abstract—A practical preparation of benzazepine derivatives using a series of radical and ionic reactions is reported. This approach was applied to the synthesis of tolvaptan, a very promising vasopressin V_2 receptor antagonist currently in clinical trials. © 2006 Elsevier Ltd. All rights reserved.

1. Introduction

Benzazepines and related compounds such as benzazepinones and benzodiazepines constitute a wide class of compounds with diverse and often important pharmacological properties. Paullones (1),¹ for example (Fig. 1), show important antitumor activity, while others behave as inhibitors of angiotensin converting enzyme² and as analgesics.³ Recent studies revealed that *N*-substituted benzazepines can act as powerful vasopressin V₂ receptor antagonists and thus have a potential for the treatment of heart diseases.⁴ Some of the most promising derivatives of this group are tolvaptan (2)⁵ and OPC-31260 (3),⁶ currently undergoing clinical trials (Fig. 1).

Even though benzazepines are well known and studied structures, difficulties arise in their preparation especially when a C5-substituent is required or if electron withdrawing groups are present in the aromatic ring. The preparation of tolvaptan, for example, required 11 linear steps, some involving harsh conditions that limit the generality of this approach (Scheme 1). Besides, even if the final steps of this route seem trivial, the yields are low, thus resulting in a very low overall yield

Keywords: Xanthate; Benzazepines; Beckmann rearrangement; Tolvaptan.

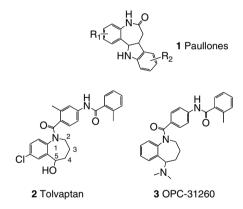


Figure 1. Some vasopressin V_2 receptor antagonists.

(<1%). In addition, the introduction of a C5-substituent other than oxygen would require extra steps in order to modify the ketone precursor. In other words, this route lacks flexibility, and cannot be easily transposed to the synthesis of analogues.

One practical method for the preparation of benzazepines is by application of the Beckmann rearrangement to tetralone ketoximes⁷ followed by amide reduction. Although this transformation is very effective, the main problem lies in the preparation of substituted tetralones. A few years ago, we reported a new method for the preparation of α -tetralones by means of a tin-free radical chemistry.⁸ This method allows the synthesis of a wide variety of substituted α -tetralones under neutral, mild conditions. More recently, this methodology was

[★] This work has been the subject of a patent. Cordero-Vargas, A.; Quiclet-Sire, B.; Zard, S. Z. Procédé utile pour la préparation de benzazépines et dérivés de celles-ci. France. French patent. FR-0450416. 02 March 2004.

^{*}Corresponding author. Tel.: +33 1693 34867; fax: +33 1693 33851; e-mail: zard@poly.polytechnique.fr

Scheme 1. Reported synthesis of tolvaptan.

employed for the preparation of substituted naphthalenes⁹ and *C*-arylglycosides¹⁰ as well as for the first total synthesis of the natural product 10-norparvulenone.¹¹ We have now found that this chemistry can be applied in an efficient practical synthesis of tolvaptan. More generally, it represents a flexible entry into benzazepines with various substitution patterns hitherto not easily accessible.

2. Results and discussion

The main general features of our route are outlined in Scheme 2. Tolvaptan (2) could derive from benzazepine 4 by acylation and deprotection of the alcohol group. The latter can be derived, in the crucial step of the synthesis, by a Beckmann rearrangement and amide reduction from ketoxime 6. The protected alcohol group has to resist the Beckmann rearrangement conditions. This is not an easily fulfilled requirement since we had shown that adducts 7 readily undergo aromatisation to naphthols upon exposure to acid. Furthermore, the lability to acid increases as the lactam 5 stage is reached because of the electron releasing effect of the nitrogen lone pair. Finally, tetralone 7, the direct precursor of 6, would be easily prepared in a convergent manner from an acetophenone xanthate (8) and an appropriate olefin (9), the latter serving as the radical trap.

Scheme 2. Retrosynthetic analysis for tolvaptan.

Our synthesis starts with the preparation of the required tetralones 11a-c as previously reported. Thus, xanthate group transfer addition of xanthates 8a-c, 12 using dilauroyl peroxide (DLP) as radical initiator and 1,2-dichloroethane (DCE) as the solvent, produces adducts 10a-c in excellent yield. The latter were then subjected to the same free radical conditions in the presence of a catalytic amount of camphorsulfonic acid (CSA)¹³ to generate tetralones 11a-c by ring closure to the aromatic ring. This step requires stoichiometric quantities of the peroxide, which acts both as initiator for the radical cyclisation and as oxidant for the intermediate cyclohexadienyl radical (Scheme 3).

The preparation of the corresponding oximes 12a–c was easily achieved by reaction of 11a–c with NH₂OH–HCl and sodium acetate in ethanol/water (Scheme 4). Having obtained the required precursors for the Beckmann rearrangement, we proceeded to prepare key intermediates 13a–c. Unfortunately, all standard conditions for mediating the ring expansion failed. Our worries concerning the lability of the benzylic ester as the main stumbling block of the synthesis were thus well founded.

Scheme 3. Radical sequence for the preparation of tetralones.

11a-c
$$\frac{NH_{2}OH.HCI}{AcONa}$$
 R $\frac{12a}{EtOH/H_{2}O}$ R $\frac{12a}{I}$ R = CI, 96% $\frac{12b}{I}$ R = F, 92% $\frac{12b}{I}$ R = OMe, 92% $\frac{13a}{I}$ R = CI, 40% (3 steps) $\frac{13a}{I}$ R = F,61% (3 steps) $\frac{13b}{I}$ R = OMe, 21% (3 steps) $\frac{13b}{I}$ R = OMe, 21% (3 steps)

Scheme 4. Beckmann rearrangement-reduction sequence.

Ultimately, after careful experimentation, we found that treatment of oximes 12a–c with an excess of PCl_5 in CH_2Cl_2 produced an α,α -dichloro lactam, ¹⁴ which was then subjected to reduction with Zn powder in acetic acid. The amide carbonyl could then be selectively reduced ¹⁵ by BH_3 –THF complex to give key benzazepines 13a–c in good overall yields, except for the methoxy substituted derivative. This is not surprising since the presence of the electron donating group increases the sensitivity of the pivalate group to the presence of acid during the Beckmann rearrangement.

Completion of the synthesis of tolvaptan is shown in Scheme 5. With the benzazepine precursor in place, the next step toward tolvaptan was the acylation of the secondary amine with 2-methyl-4-nitro benzoyl chloride. The high efficiency of this seemingly trivial step contrasts with the analogous acylation described

13a-b
$$\frac{O}{NEt_3/DCM}$$
 $\frac{NO_2}{rt}$ $\frac{NO_2}{NEt_3/DCM}$ $\frac{14a}{14b}$, $R=Cl$, 98% $\frac{14b}{14b}$, $R=F$, 97% $\frac{SnCl_2}{EtOH/HCl}$ $\frac{SnCl_2}{rt}$ $\frac{EtOH/HCl}{rt}$ $\frac{NH_2}{NEt_3/DCM}$ $\frac{NEt_3/DCM}{rt}$ $\frac{NH_2}{NEtOH/EtOH}$ $\frac{NH_2}{NH_2}$ $\frac{NH_2}{$

Scheme 5. Synthesis of tolvaptan and its fluoro analogue.

by the Otsuka chemists (Scheme 1). In their route, the presence of the electron withdrawing ketone function deactivates the nitrogen and lowers considerably the efficiency of the acylation step. Reduction of the nitro group of 14a-b with tin (II) chloride in ethanol/HCl afforded anilines 15a-b. Finally, acylation of the latter with 2-methyl benzoyl chloride and saponification of the ester group under basic conditions allowed us to obtain tolvaptan (2) and its 5'-fluoro analogue (16) in excellent yield.

Our method for the synthesis of α-tetralones is quite flexible and several substituents can be introduced into the molecule just by choosing the appropriate radical trap. In order to show the scope and applicability of this method, we prepared some analogues possessing a hydrocarbon chain at C5 (Scheme 6). Thus, reaction of xanthate **8b** with a small amount of DLP in the presence of allyl cyanide afforded adduct **17** in 81% yield. Tetralone **18** was prepared from **17** in 44% yield after the CSA catalyzed cyclisation step. The final three-step sequence was then applied to oxime **19**, affording benzazepine **20** in good overall yield.

As seen in Scheme 6, the presence of the nitrile group in **20** is not arbitrary. This functional group is highly useful because it could be used as a starting point to prepare a series of other benzazepines possessing a primary amine **(21)**, an acid **(22)**, 4b or an oxathiadiazole oxide **(23)**. 17 The latter structure also exhibits interesting antihyperglycemic activity. 15

While α -halo lactams have been reported as important intermediates for the synthesis of C3-substituted

Scheme 6. Synthesis of further analogues.

Scheme 7. Trapping the intermediate in the Beckmann rearrangement.

Scheme 8. Synthesis of a dichlorobenzazepine.

benzazepinones,² benzazepines bearing this substitution pattern are rare. It is known⁷ that the intermediate of the Beckmann reaction is an iminium ion (24) that can be trapped by a nucleophile in order to generate the corresponding imines (25) (Scheme 7).

We exploited this feature in order to prepare a novel benzazepine derivative possessing a halide in the 3-position, which would be difficult to access otherwise. When oxime 12a was treated with an excess of PCl₅ in CH₂Cl₂ and quenched with a suspension of NaBH₄ in ethanol, compound 26 was obtained in 48% overall yield (Scheme 8). This molecule could, of course, be used as starting material for the synthesis of yet another tolvaptan analogue.

3. Conclusion

In summary, we have illustrated a useful application of the xanthate methodology. The mild conditions of this protocol allow the preparation of a great diversity of functionalized benzazepines tedious to obtain by more traditional routes. The efficiency of our approach to tolvaptan (2) is indicated by the good overall yield (21%), which contrasts with the low overall yield reported for the original route (<1%). In addition, the synthesis of the 5'-fluoro derivative 16 (20.6% overall yield) as well as the preparation of analogues 20 and 26 underscores its flexibility.

4. Experimental

4.1. General considerations

All reactions were carried out under an inert atmosphere. Commercial reagents were used as received without further purification. All products were purified by using silica gel SDS 60 C.C. 40–63 or by crystallisation. NMR spectra were recorded in CDCl₃ with TMS as an internal standard at room temperature on a Bruker AMX400 operating at 400 MHz for ¹H and 100 MHz for ¹³C. Infrared Absorption spectra were recorded as a solution in CCl₄ with a Perkin-Elmer 1600 Fourier

Transform Spectrophotometer. Mass spectra were recorded with a HP 5989B mass spectrometer. Melting points were determined by Reichert microscope apparatus and were uncorrected.

4.1.1. 2,2-Dimethyl-propionic acid 1-ethoxythiocarbonylsulfanyl-4-(4-methoxy-phenyl)-4-oxo-butyl ester (10c). A solution of **8c** (0.5 g, 1.85 mmol) and of 0.55 mL (2.7 mmol) of vinyl pivalate in 1,2-dichloroethane (2 mL) was refluxed for 15 min under argon. Lauroyl peroxide (DLP) was then added (5 mol %) to the refluxing solution, followed by additional portions (2 mol % every 90 min). When starting material was totally consumed (after addition of 7.5 mol % of DLP), the crude mixture was cooled to room temperature, concentrated under reduced pressure and purified by flash column chromatography (silica gel, petroleum ether-ethyl acetate, 95:5) to give **10c** as a yellow oil (86% yield): ¹H NMR (CDCl₃, 400 MHz) δ 7.92 (d, 2H, CH arom, J = 8.8 Hz), 6.93 (d, 2H, CH arom, J = 8.8 Hz), 6.71 (t, 1H, CH-S, J = 8.0 Hz), 4.65–4.59 (m, 2H, O-C H_2), 3.87 (s, 3H, OC H_3), 3.08 (dq, 2H, CO–C H_2 , J = 7.2, 4.4 Hz), 2.42–2.35 (m, 2H, $CH-CH_2$), 1.41 (t, 3H, CH_2-CH_3 , J = 6.0 Hz), 1.19 (s, 9H, $(\overline{CH_3})_3$); ¹³C NMR $(CDCl_3, 100 \text{ MHz}) \delta 210.3 (CS), 196.6 (CO), 176.9$ (O-CO), 163.7 (C-OMe), 130.9 (C-CO), 130.4 (CH arom), 113.9 (CH arom), 80.4 (CH-S), 70.4 (O-CH₂), 55.6 (OCH₃), 43.5 (C-(CH₃)₃), 33.9 (CO-CH₂), 28.8 (CH-CH₂), 27.1 (3C, (CH₃)₃), 13.8 (CH₂-CH₃); MS $(CI+NH_3)$ m/z: 416 (MH^++NH_3) , 399 (MH^+) , 297 $(MH^+-PivOH)$; IR (cm^{-1}, CCl_4) : 1738 (O-C=O), 1683 (C=O), 1229 (C=S).

4.1.2. Dithiocarbonic acid [1-cyanomethyl-4-(4-fluorophenyl)-4-oxo-butyl ester ethyl ester (17). A solution of **8b** (2.0 g, 7.74 mmol) and of 1.25 mL (1.03 g, 15.48 mmol) of allyl cyanide in 1,2-dichloroethane (8 mL) was refluxed for 15 min under argon. Lauroyl peroxide (DLP) was then added (5 mol %) to the refluxing solution, followed by additional portions (2 mol % every 90 min). When starting material was totally consumed (after addition of 15 mol % of DLP), the crude mixture was cooled to room temperature, concentrated under reduced pressure and purified by flash column chromatography (silica gel, petroleum ether-ethyl acetate, 9:1) to give 17 as a yellow oil (81% yield): ${}^{1}H$ NMR (CDCl₃, 400 MHz) δ 7.98 (dd, 2H, CH arom, J = 8.4, 5.2 Hz), 7.14 (t, 2H, CH arom, J = 8.4 Hz, 4.63 (m, 2H, O-C H_2), 4.01 (dddd, 1H, C*H*–S, J = 15.2, 5.3, 5.3, 5.3 Hz), 3.19 (t, 2H, CO–C H_2 , J = 7.2 Hz), 2.96 (t, 2H, C H_2 – CN), 2.39 (ddt, 1H, CO-CH₂-CH₂, J = 10.9, 7.2, 4.3 Hz), 2.15 (dddd, 1H, CO–CH₂–C H_2 , J = 18.0, 7.2, 6.8, 6.8 Hz), 1.42 (t, 3H, CH₂–C H_3 , J = 7.0 Hz); 7.2, 6.8, 6.8 Hz), 1.42 (t, 3H, CH₂=CH₃, $\delta = 7.0$ Hz), 13C NMR (CDCl₃, 100 MHz) δ 212.0 (CS), 196.5 (CO), 165.9 (d, 1C, C–F, $^{1}J_{C-F} = 255.2$ Hz), 132.9 (C–CO), 130.7 (d, 2C, CH arom, $^{2}J_{C-F} = 13.0$ Hz), 117.1 (C=N), 115.9 (d, 2C, CH arom, $^{3}J_{C-F} = 255.2$ (CH arom, $^{3}J_{C-F} = 255.2$ 22.0 Hz), 70.7 (O–*C*H₂), 46.2 (*C*H–S), 35.4 (CO–*C*H₂), 26.7 (*C*H₂–CN), 24.5 (CO–*C*H₂–*C*H₂), 13.8 $(CH_2-CH_3); MS (CI+NH_3) m/z: 343 (MH^++NH_3),$ 326 (MH $^+$), 205 (MH $^+$ –SC(S)OEt); IR (cm $^{-1}$, CCl₄): 2250 (C=N), 1741 (C=O), 1236 (C=S).

- 4.1.3. 2,2-Dimethyl-propionic acid 7-methoxy-4-oxo-1,2,3,4-tetrahydro-naphthalen-1-yl ester (11c). A solution of **10c** (3.0 g, 7.50 mmol) in 1,2-dichloroethane (75 mL) was refluxed for 15 min under argon. Laurovl peroxide (DLP) was then added portionwise (20 mol % per hour) to the refluxing solution. When starting material was totally consumed (after addition of 1.2 equiv DLP), the crude mixture was cooled to room temperature, concentrated under reduced pressure and purified by flash column chromatography (silica gel, petroleum ether-AcOEt, 9:1 to 8:2) and recrystallised with ethanol to give tetralone 11c (30% yield) as white crystals (mp 80 °C): ¹H NMR (CDCl₃, 400 MHz) δ 8.06 (d, 1H, CH arom, J = 8.0 Hz), 6.96 (dd, 1H, CH arom, J = 8.0, 4.0 Hz), 6.89 (d, 1H, CH arom, J = 4.0 Hz), 6.08 (dd, 1H, CH– OPiv, J = 8.0, 4.0 Hz), 3.89 (s, 3H, OC H_3), 2.87 (ddd, 1H, CO-C H_2 , J = 18.0, 9.0, 6.0 Hz), 2.66 (ddd, 1H, $CO-CH_2$, J = 16.0, 8.0, 4.0 Hz), 2.45–2.37 (m, 1H, $CO-CH_2-CH_2$), 2.29–2.21 (m, 1H, $CO-CH_2-CH_2$). 1.26 (s, 9H, $(CH_3)_3$); ¹³C NMR (CDCl₃, 100, 5 MHz) δ 195.8 (CO), 177.9 (O-CO), 164.0 (C-OMe), 143.7 (C-CO), 129.8 (CH arom), 125.5 (C-C-CO), 114.9 (CH arom), 111.9 (CH arom), 69.1 (CH-OPiv), 55.6 (OCH_3) , 39.1 $(C-(CH_3)_3)$, 34.6 $(CO-CH_2)$, 28.8 $(CO-CH_3)$ CH_2-CH_2), 27.18 (3C, $(CH_3)_3$); MS (CI+NH₃) m/z: 294 (MH⁺+NH₃), 277 (MH⁺), 176 (MH⁺-PivOH); IR (cm⁻¹, CCl₄): 1731 (O-C=O), 1687 (C=O), 1146 (O-C=O); Anal. calcd. for $C_{16}H_{20}O_4$: C, 69.54; H, 7.3. Found: C, 69.07; H, 7.27.
- 7-Fluoro-4-oxo-1,2,3,4-tetrahydro-naphthalen-1yl)-acetonitrile (18). A solution of 17 (2.0 g, 6.14 mmol) and of CSA (0.14 g, 0.61 mmol) in 1,2-dichloroethane (61 mL) was refluxed for 15 min under argon. Lauroyl peroxide (DLP) was then added portionwise (20 mol % per hour) to the refluxing solution. When starting material was totally consumed (after addition of 1.2 equiv DLP), the crude mixture was cooled to room temperature, concentrated under reduced pressure and purified by flash column chromatography (silica gel, petroleum ether-AcOEt, 8:2) and recrystallised with petroleum ether to give tetralone 18 (44% yield) as a yellow solid (mp 126–128 °C): 1 H NMR (CDCl₃, 400 MHz) δ 8.12 (dd, 1H, CH arom, J = 9.0, 5.8 Hz), 7.1 (dt, 1H, CH arom, J = 8.2, 2.4 Hz), 7.05 (d, 1H, CH arom, J = 9.6 Hz), 3.38 (tt, 1H, CH, J = 11.6, 6.1 Hz), 2.75– 2.83 (m, 3H, CO– CH_2 and CH_2 –CN), 2.68 (ddd, 1H, CH_2 -CN, J = 18.0, 7.4, 5.0 Hz), 2.45 (ddd, 1H, CO- CH_2-CH_2 , J = 19.0, 9.4, 4.6 Hz), 2.20–2.28 (m, 1H, CO–CH₂–CH₂); 13 C NMR (CDCl₃, 100 MHz) δ 194.9 (CO), 166.06 (d, 1C, C-F, ${}^{1}J_{\text{C-F}} = 256 \text{ Hz}$), 146.1 (C-CO), 131.3 (d, 1C, CH arom, ${}^{3}J_{\text{C-F}} = 10.9 \text{ Hz}$), 122.0 (C \equiv N), 117.6 (*C*-CCO), 115.9 (d, 1C, *CH* arom, $^2J_{C-F}$ = 23 Hz), 114.2 (d, 1C, *CH* arom, $^2J_{C-F}$ = 22 Hz), 35.1 (*CH*), 34.9 (CO-*CH*₂), 27.6 (*CH*₂-CN), 22.9 (CO- CH_2-CH_2); MS (CI+NH₃) m/z: 221 (MH⁺+NH₃), 204 (MH^+) ; IR (cm^{-1}, CCl_4) : 2254 $(C \equiv N)$, 1693 $(C \equiv O)$, 1250 (C-F); Anal. calcd. for C₁₂H₁₀NOF: C, 70.93; H, 4.96; N, 6.44. Found: C, 70.53; H, 5.03; N, 6.56.
- **4.1.5.** General procedure for the preparation of oximes. To a stirred solution of the corresponding tetralone (1 mmol) in ethanol (0.75 mL) was added a solution of

hydroxylamine hydrochloride (1.3 mmol) and anhyd sodium acetate (1.2 mmol) in 0.3 mL of water. The resulting mixture was then refluxed for 2 h. After consumption of all the starting material, ethanol was evaporated and the resulting mixture was diluted with water, extracted with ethyl acetate, dried over Na₂SO₄ and concentrated. The residue was purified by crystallisation to afford the desired oximes.

- 4.1.6. 2,2-Dimethyl-propionic acid 4-[(E)-hydroxyimino]-7-chloro-1,2,3,4-tetrahydro-naphthalen-1-yl ester (12a). According to the general procedure, a solution of 1.99 g (7.11 mmol) of tetralone 11a in 5.3 mL of ethanol is treated with a solution of NH₂OH · HCl (0.6 g, 9.24 mmol) and anhyd sodium acetate (1.16 g, 8.53 mmol) in water (2.1 mL) to furnish oxime 12a as a yellow solid (96%), mp 110-112 °C (petroleum ether): ¹H NMR (CDCl₃, 400 MHz) δ 9.19 (br. 1H, OH), 7.86 (d, 1H, CH arom, J = 8.0 Hz), 7.35 (d, 1H, CH arom, J = 4.0 Hz), 7.29 (dd, 1H, CH arom, J = 8.0, 4.0 Hz), 5.89 (t, 1H, CH– OPiv, J = 4.0 Hz, 2.93 (t, 2H, C(NOH)–C H_2 , J = 6.0 Hz), 2.07 (m, 2H, CH-C H_2), 1.22 (s, 9H, $(CH_3)_3$); ¹³C NMR (CDCl₃, 100 MHz) δ 178.0 (O–CO), 153.3 (C-NOH), 138.3 (C-C(NOH)), 135.6 (C-Cl), 129.0 (CH arom), 128.9 (C-CH), 128.0 (CH arom), 125.7 (CH arom), 69.0 (CH–OPiv), 39.1 (C–(CH₃)₃), 27.2 (3C, (CH₃)₃), 26.3 (C(NOH)–CH₂), 19.3 (CH–CH₂).
- 4.1.7. 2,2-Dimethyl-propionic acid 4-[(E)-hydroxyimino]-7-fluoro-1,2,3,4-tetrahydro-naphthalen-1-yl ester (12b). According to the general procedure, a solution of 1.14 g (4.31 mmol) of tetralone 11b in 3.2 mL of ethanol is treated with a solution of NH₂OH·HCl (0.36 g, 5.6 mmol) and anhyd sodium acetate (0.43 g, 5.17 mmol) in water (1.3 mL) to furnish oxime 12b as a yellow solid (92%), mp 125-128 °C (petroleum ether): ${}^{1}H$ NMR (CDCl₃, 400 MHz) δ 9.10 (br, 1H, OH), 7.92 (dd, 1H, CH arom, J = 8.8, 6.0 Hz), 7.05 (dt, 1H, CH arom, J = 8.5, 2.5 Hz), 7.02 (dd, 1H, CH arom, J = 8.2, 3.0 Hz), 5.89 (dd, 1H, CH-OPiv, J = 7.0, 3.8 Hz), 2.94 (m, 2H, C(NOH)-C H_2), 2.07 (m, 2H, CH-C H_2), 1.23 (s, 9H, C H_3); ¹³C NMR (CDCl₃, 100 MHz) δ 178.0 (O–CO), 164.7 (C–NOH), 163.5 (d, 1C, C–F, ${}^{1}J_{C}$ –F = 250 Hz 153.3 (C–C–(NOH)), 139.2 (d, 1C, CH arom, ${}^{3}J_{C-F} = 8.0 \text{ Hz}$), 126.5 (C–CH), 116.2 (d, 1C, CH arom, ${}^{2}J_{C-F} = 22 \text{ Hz}$), 114.3 (d, 1C, CH arom, ${}^{2}J_{C-F} = 23 \text{ Hz}$), 69.1 (CH–OPiv), 39.1 $(C-(CH_3)_3)$, 27.2 (3C, $(CH_3)_3$), 26.4 (C(NOH)- CH_2), 19.5 (CH–*C*H₂); MS (CI+NH₃) *m/z*: 297 (MH⁺+NH₃), 280 (MH⁺), 179 (MH⁺–OPiv); IR (cm⁻¹, CCl₄): 3593 (N-OH), 1730 (O-C=O), 1279 (O-C=O).
- **4.1.8. 2,2-Dimethyl-propionic acid 4-[(E)-hydroxyimino]7-methoxy-1,2,3,4-tetrahydro-naphthalen-1-yl ester (12c).** According to the general procedure, a solution of 2.2 g (0.78 mmol) of tetralone **11c** in 0.6 mL of ethanol is treated with a solution of NH₂OH·HCl (0.07 g, 1.0 mmol) and anhyd sodium acetate (1.13 g, 0.93 mmol) in water (0.2 mL) to furnish oxime **12c** as a yellow solid (92%), mp 114–115 °C (petroleum ether): ¹H NMR (CDCl₃, 400 MHz) δ 9.34 (br, 1H, O*H*), 7.86 (d, 1H, C*H* arom, J = 8.0 Hz), 6.90 (d, 1H, C*H* arom, J = 2.0 Hz), 6.87 (dd, 1H, C*H* arom, J = 7.8, 2.2 Hz),

5.91 (dd, 1H, CH–OPiv, J = 6.0, 4.0 Hz), 3.82 (s, 3H, OC H_3), 2.93 (t, 2H, C(NOH)– CH_2 , J = 6.8 Hz), 2.04–2.11 (m, 2H, CH– CH_2), 1.22 (s, 9H, (C H_3)₃); ¹³C NMR (CDCl₃, 100 MHz) δ 178.13 (O–CO), 160.7 (C–OMe), 153.8 (C–NOH), 138.4 (C–C(NOH)), 125.8 (CH arom), 123.0 (C–CH), 115.4 (CH arom), 112.2 (CH arom), 69.6 (CH–OPiv), 55.4 (OCH₃), 39.1 (C–(CH₃)₃), 27.2 (3C, (CH₃)₃), 26.6 (C(NOH)–CH₂), 19.4 (CH–CH₂); MS (CI+NH₃) m/z: 292 (MH⁺), 191 (MH⁺–OPiv); IR (cm⁻¹, CCl₄): 3596 (N–OH), 1727 (O–C=O), 1604 (C=N–OH), 1272 (O–C=O).

4.1.9. {7-Fluoro-4-[(E)-hydroxyimino]-1,2,3,4-tetrahydronaphthalen-1-yl}-acetonitrile (19). According to the general procedure, a solution of 0.4 g (1.96 mmol) of tetralone 18 in 0.9 mL of ethanol is treated with a solution of NH₂OH·HCl (0.16 g, 2.55 mmol) and anhyd sodium acetate (0.32 g, 2.36 mmol) in water (0.4 mL) to furnish oxime 19 as a yellow solid (78%), mp 167– 168 °C (ethyl acetate/petroleum ether): 1H NMR $(CDCl_3/DMSO-d_6, 400 MHz) \delta 9,94 (br, 1H, OH),$ 7.97 (dd, 1H, CH arom, J = 8.8, 6.0 Hz), 6.93–7.0 (m, 2H, CH arom), 3.16-3.22 (m, 1H, CH), 2.96 (dt, 1H, C(NOH)– CH_2 , J = 18.8, 5.2 Hz), 2.73 (m, 1H, C(NOH)– CH_2), 2.58–2.66 (m, 1H, CH_2 –CN), 2.05– 2.12 (m, 2H, CH–C H_2); ¹³C NMR (CDCl₃, 100 MHz) δ 162.3 (d, 1C, C–F, ¹ J_{C –F = 250 Hz), 150.7 (C=NOH), 140.0 (*C*–C=NOH), 126.8 (*C*=N), 126.2 (d, 1C, *C*H arom, ${}^3J_{\text{C-F}} = 7.7 \text{ Hz}$), 117.8 (*C*–CH), 114.4 (d, 1C, *C*H arom, ${}^2J_{\text{C-F}} = 15.0 \text{ Hz}$), 113.6 (d, 1C, *C*H arom, $^{2}J_{C-F}$ = 23 Hz), 34.7 (*CH*), 24.8 (C(NOH)–*CH*₂), 21.9 (CH_2-CN) , 19.1 $(CH-CH_2)$; MS $(CI+NH_3)$ m/z: 236 $(MH^{+}+NH_{3})$, 219 (MH^{+}) ; IR (cm^{-1}, CCl_{4}) : 3591 (N-OH), 1741 (C=N-OH), 1239 (O-C=O).

4.1.10. General procedure for the preparation of benzazepines. To a stirred suspension of PCl₅ (4 mmol) in dry CH₂Cl₂ (10 mL) at 0 °C was slowly added a solution of the corresponding oxime (1 mmol) in 10 mL of dry CH₂Cl₂. The mixture was stirred at room temperature for 2 h, neutralised with saturated aqueous NaHCO₃, extracted with CH₂Cl₂, dried over NaSO₄ and concentrated under reduced pressure. The residue was then dissolved in 10 mL of acetic acid and heated to reflux. Then, powdered Zn (6 mmol) was slowly added to the reaction mixture. The resulting mixture was then refluxed for further 30 min. When starting material was totally consumed, the crude mixture was diluted with ethyl acetate, filtered through Celite, washed with saturated aqueous NaHCO3 and concentrated in vacuo. The residue was then dissolved in THF (1.5 mL) and added dropwise to a solution of BH₃·THF (2 mmol) in THF (1.5 mL) at 0°C. The resulting solution was then refluxed for 30 min, acidified with saturated aqueous citric acid and the solvent was evaporated under reduced pressure. The residue was then basified with saturated aqueous Na₂CO₃, extracted with CH₂Cl₂, dried over Na₂SO₄ and concentrated under reduced pressure. The title compound is purified by flash chromatography.

4.1.11. 2,2-Dimethyl-propionic acid 7-chloro-2,3,4,5-tet-rahydro-1*H***-benzo**[*b*]**azepin-5-yl ester (13a).** Following the general method, this compound was obtained from

oxime 12a as a white solid (40% over 3 steps), mp 65–66 °C (petroleum ether): ¹H NMR (CDCl₃, 400 MHz) δ 7.27 (d, 1H, CH arom, J = 2.4 Hz), 7.02 (dd, 1H, CH arom, J = 8.4, 2.4 Hz), 6.64 (d, 1H, CH arom, J = 8.8 Hz), 5.84 (d, 1H, CH-OPiv, J = 8.0 Hz), 3.83 (br, 1H, NH), 3.25 (dt, 1H, NH– CH_2 , J = 13.2, 4.2 Hz), 2.93 (ddd, 1H, NH–C H_2 , J = 13.2, 9.6, 3.2 Hz), 1.80–2.0 (m, 4H, CH– CH_2 and NH– CH_2 – CH_2), 1.27 (s, 9H, $(CH_3)_3$); ^{13}C NMR $(CDCl_3)_3$ 100 MHz) δ 177.3 (O–CO), 147.5 (C–NH), 131.5 (C-Cl), 128.1 (CH arom), 127.7 (CH arom), 125.2 (C-CH), 120.8 (CH arom), 73.7 (CH-OPiv), 47.5 $(NH-CH_2)$, 41.7 $(C-(CH_3)_3)$, 31.3 $(CH-CH_2)$, 27.3 $(3C, (CH_3)_3), 27.0 (NH-CH_2-CH_2); MS (CI+NH_3)$ m/z: 284 (MH⁺), 282 (MH⁺), 184 (MH⁺–PivOH) 181 (MH⁺–PivOH); IR (cm⁻¹, CCl₄): 3386 (NH), 1729 (O-C=O), 1151 (O-C=O); Anal. calcd. $C_{15}H_{20}NO_2Cl$: C, 63.94; H, 7.15. Found: C, 63.75; H, 7.15.

4.1.12. 2.2-Dimethyl-propionic acid 7-fluoro-2.3.4.5-tetrahydro-1*H*-benzo[*b*]azepin-5-yl ester (13b). Following the general method, this compound was obtained from oxime 12b as a white solid (61% over 3 steps), mp 58-60 °C (petroleum ether): ¹H NMR (CDCl₃, 400 MHz) δ 7.05 (dd, 1H, CH arom, J = 9.4, 3.0 Hz), 6.78 (td, 1H, CH arom, J = 8.3, 3.1 Hz), 6.68 (dd, 1H, CH arom, J = 8.4, 4.8 Hz), 5.84 (d, 1H, CH–OPiv, J = 10.0 Hz), 3.58 (br, 1H, NH), 3.26 (dt, 1H, NH– CH_2 , J = 13.2, 4.2 Hz), 2.79 (td, 1H, NH– CH_2 , J = 11.7, 2.8 Hz), 1.7-2.0 (m, 4H, CH-CH₂ and NH-CH₂-CH₂), 1.26 (s, 9H, $(CH_3)_3$); ¹³C NMR (CDCl₃, 100 MHz) δ 177.2 (O-CO), 157.7 (d, 1C, C-F, $^{1}J_{C-F} = 236 \text{ Hz}$), 144.4 (C-NH), 133.2 (C-CH), 120.9 (d, 1C, CH arom, ${}^2J_{\text{C-F}} = 7.0 \text{ Hz}$), 114.1 (d, 1C, CH arom, ${}^2J_{\text{C-F}} = 7.0 \text{ Hz}$), 113.7 (d, 1C, CH arom, ${}^3J_{\text{C-F}} = 24 \text{ Hz}$), 73.6 (CH-OPiv), 47.6 (NH-CH₂), 39.0 (C-(CH₃)₃), 32.0 $(NH-CH_2-CH_2)$, 27.5 $(CH-CH_2)$, 27.3 $(3C, (CH_3)_3)$; MS (CI+NH₃) m/z: 266 (MH⁺), 165 (MH⁺-OPiv); IR (cm^{-1}, CCl_4) : 3385 (NH), 1729 (O-C=O), 1152 (O-C=O); Anal. calcd. for $C_{15}H_{20}NO_2F$: C, 67.9; H, 7.6. Found: C, 67.93; H, 7.62.

4.1.13. 2,2-Dimethyl-propionic acid 7-methoxy-2,3,4,5tetrahydro-1H-benzo[b]azepin-5-yl ester (13c). According to the general method, this compound was obtained from oxime 12c as a yellow oil (21% over 3 steps): ¹H NMR (CDCl₃, 400 MHz) δ 6.93 (d, 1H, CH arom, J = 2.4 Hz), 6.66–6.7 (m, 2H, CH arom), 5.87 (d, 1H, CH-OPiv, J = 9.2 Hz), 3.76 (s, 3H, OC H_3), 3.24 (dt, 1H, NH-C H_2 , J = 12.8 et 4.2 Hz), 2.78 (td, 1H, NH- CH_2 , J = 11.7, 2.8 Hz), 1.75–2.0 (m, 4H, CH–C H_2 and NH-CH₂-CH₂), 1.28 (s, 9H, (CH₃)₃); 13 C NMR (CDCl₃, 100 MHz) δ 189.1 (O–CO), 141.9 (C–OMe), 134.0 (C– NH), 130.2 (C-CH), 121.0 (CH arom), 112.9 (CH arom), 112.5 (CH arom), 74.1 (CH-OPiv), 55.6 (OCH₃), 48.2 $(NH-CH_2)$, 39.1 $(C-(CH_3)_3)$, 32.3 $(NH-CH_2-CH_2)$, 27.7 (CH–CH₂), 27.4 (3C, (CH₃)₃); MS (CI+NH₃) m/z: 278 (MH⁺), 177 (MH⁺–PivOH); IR (cm⁻¹, CCl₄): 3450 (NH), 1727 (O-C=O), 1156 (O-C=O).

4.1.14. (7-Fluoro-2,3,4,5-tetrahydro-1*H*-benzo[*b*]azepin-5-yl)-acetonitrile (20). According to the general procedure, this compound was obtained from oxime 19 as a white solid

(39% over 3 steps), mp 74–75 °C (petroleum ether): ¹H NMR (CDCl₃, 400 MHz) δ 6.84 (dd, 1H, CH arom, J = 9.2, 2.4 Hz, 6.78 (td, 1H, CH arom, J = 8.2, 3.0 Hz), 6.67 (dd, 1H, CH arom, J = 8.4, 4.8 Hz), 3.59 (sl, 1H, NH), 3.19–3.29 (m, 2H, CH and NH–CH₂), 3.0 (dd, 1H, CH_2 -CN, J = 16.6, 8.6 Hz), 2.81 (dd, 1H, CH_2 -CN, J = 16.6, 7.4 Hz), 2.72 (ddd, 1H, NH–C H_2 , J = 12.6, 10.8, 2.0 Hz), 2.05-2.1 (m, 1H, NH-CH₂-CH₂), 1.84-1.95 (m, 1H, NH-CH₂-CH₂), 1.71-1.79 (m, 2H, CH- CH_2); ¹³C NMR (CDCl₃, 100 MHz) δ 157.8 (d, 1C, C-F, $J_{\text{C-F}} = 241 \text{ Hz}$), 145.7 (*C*-NH), 134.4 (d, 1C, *C*H arom, ${}^{3}J_{C-F} = 241 \text{ Hz}$), 143.7 (C=N1), 134.7 (d, 1c, cH arom, ${}^{3}J_{C-F} = 6.0 \text{ Hz}$), 121.8 (C=N), 119.2 (C-CH), 116.7 (d, 1C, CH arom, ${}^{2}J_{C-F} = 22.1 \text{ Hz}$), 114.2 (d, 1C, CH arom, ${}^{2}J_{C-F} = 19.1 \text{ Hz}$), 48.9 (NH-CH₂), 42.5 (CH), 29.9 (CH₂-CN), 26.1 (NH-CH₂-CH₂), 19.1 (CH-CH₂); MS $(CI+NH_3) m/z$: 205 (MH^+) ; IR (cm^{-1}, CCl_4) : 3384 (NH), 2246 (C \equiv N), 1253 (*O*–*C* \equiv O); Anal. calcd. for C₁₂H₁₃ N₂F: C, 70.57; H, 6.42. Found: C, 70.42; H, 6.55.

2.2-Dimethyl-propionic acid 3,3,7-trichloro-2,3,4,5-tetrahydro-1*H*-benzo[*b*]azepin-5-yl ester (26). To a stirred suspension of PCl₅ (0.28 g, 1.35 mmol) in dry CH₂Cl₂ (1,5 mL) at 0 °C was slowly added a solution of oxime 12a (0.1 g, 0.34 mmol) in 1.5 mL of dry CH₂Cl₂. The mixture was stirred at room temperature for 2 h. The solution was then cooled at 0 °C and a suspension of NaBH₄ (0.13 g, 3.38 mmol) in ethanol (0.3 mL) was added dropwise. When starting material was completely consumed, water was added and the reaction mixture was extracted with CH₂Cl₂, dried over Na²SO₄ and concentrated under reduced pressure. The residue was purified by flash column chromatography (silica gel, petroleum ether-AcOEt, 95:5) and recrystallised with petroleum ether to give benzazepine 20 (48% yield over 2 steps) as a white solid (mp 103-104 °C): ¹H NMR (CDCl₃, 400 MHz) δ 7.21 (d, 1H, CH arom, J = 2.0 Hz), 7.10 (dd, 1H, CH arom, J = 8.8, 2.4 Hz), 6.72 (d, 1H, CH arom, J = 8.4 Hz), 6.03 (t, 1H, CH– OPiv, J = 5.6 Hz), 4.22 (br, 1H, NH), 3.75 (dd, 1H, $CH-CH_2$, J = 14.8, 6.8 Hz), 3.57 (d, 1H, $CH-CH_2$, J = 14.0 Hz), 3.86 (d, 2H, NH–C H_2 , J = 4.0 Hz), 1.29 (s, 9H, $(CH_3)_3$); ¹³C NMR (CDCl₃, 100 MHz) δ 177.0 (O-CO), 145.2 (C-NH), 142.3 (C-Cl), 128.5 (CH arom), 128.2 (CH arom), 126.6 (C-CH), 121.0 (CH arom), 88.4 (CCl₂), 69.2 (CH-OPiv), 61.6 (NH-CH₂), 50.1 (CH-CH $_2$), 39.0 (C-(CH $_3$) $_3$), 27.3 (3C, (CH $_3$) $_3$); MS (CI+NH₃) m/z: 352 (MH⁺), 350 (MH⁺), 250 $(MH^{+}-PivOH)$, 248 $(MH^{+}-PivOH)$; IR (cm^{-1}, CCl_{4}) : 3446 (NH), 1735 (O-C=O), 1139 (O-C=O); Anal. calcd. for C₁₅H₁₈NO₂Cl₃:C, 51.38; H, 5.17. Found: C, 51.31; H, 5.16.

4.1.16. 2,2-Dimethyl-propionic acid 7-chloro-1-(2-methyl-4-nitro-benzoyl)-2,3,4,5-tetrahydro-1*H***-benzo|***b***|azepin-5-yl ester (14a).** A solution of 2-methyl-4-nitrobenzoyl chloride (0.21 g, 1.06 mmol) in CH₂Cl₂ (2 mL) was added dropwise to an ice-cooled solution of benzazepine **13a** (0.1 g, 0.35 mmol) and triethylamine (2 mL, 0.14 g, 1.41 mmol) in CH₂Cl₂. The mixture was stirred at room temperature for 1 h, neutralised with saturated aqueous Na₂CO₃, extracted with ethyl acetate, dried over Na₂SO₄ and concentrated under reduced pressure. The residue was purified by flash column chromatography

(silica gel, petroleum ether-AcOEt, 9:1) and recrystallised with petroleum ether to give benzazepine 20 (98%) yield) as a white solid (mp 58–60 °C): ¹H NMR (CDCl₃, 400 MHz) δ 7.92 (d,1H, CH arom, J = 2.0 Hz), 7.77 (dd, 1H, CH arom, J = 8.2, 2.2 Hz), 7.15 (d, 1H, CH arom, J = 8.4 Hz), 7.14 (s, 1H, CH arom), 6.85 (dd, 1H, CH arom, J = 8.0, 2.4 Hz), 6.51 (d, 1H, CH arom, J = 8.4 Hz), 5.95 (dd, 1H, CH-OPiv, J = 5.6, 2.6 Hz), 4.74 (dt, 1H, N-C H_2 , J = 14.0, 4.1 Hz), 2.81 (ddd, 1H, N-C H_2 , J = 12.0, 10.1, 2.1 Hz), 2.49 (s, 3H, Ar-C H_3), 2.09–2.19 (m, 2H, N–CH₂–C H_2), 1.67–1.82 (m, 2H, CH–C H_2), 1.29 (s, 9H, C H_3); ¹³C NMR (CDCl₃, 100 MHz) δ 177.2 (O–CO), 167.9 (N–CO), 147.7 (C– NO₂), 141.8 (C-N), 140.2 (C-CO), 137.6 (C-Cl), 137.4 (C-CH), 134.28 (C-CH₃), 128.9 (CH arom), 128.0 (CH arom), 127.6 (CH arom), 125.3 (CH arom), 124.6 (CH arom), 121.0 (CH arom), 71.8 (CH–OPiv), 46.6 $(N-CH_2)$, 39.14 $(C-(CH_3)_3)$, 32.0 $(N-CH_2-CH_2)$, 27.3 $(3C, (CH_3)_3), 25.4 (CH-CH_2), 20.2 (Ar-CH_3); MS$ $(CI+NH_3)$ m/z: 465 (MH^++NH_3) , 463 (MH^++NH_3) , 447 (MH⁺), 445 (MH⁺), 346 (MH⁺–PivOH), 344 $(MH^{+}-PivOH)$; IR (cm^{-1}, CCl_{4}) : 1735 (O-C=O), 1659 (N-C=0), 1529 (NO_2) , 1139 (O-C=0).

4.1.17. 2,2-Dimethyl-propionic acid 7-fluoro-1-(2-methyl-4-nitro-benzoyl)-2,3,4,5-tetrahydro-1*H*-benzo[*b*]azepin-5yl ester (14b). Using the same procedure as 14a, benzazepine **14b** (97%) was obtained from **13b** (0.05 g, 0.18 mmol) as a yellow oil: ¹H NMR (CDCl₃, 400 MHz) δ 7.98 (d, 1H, CH arom, J = 2.4 Hz), 7.82 (dd, 1H, CH arom, J = 8.6, 2.6 Hz), 7.22 (d, 1H, CH arom, J = 8.8 Hz), 6.94 (d, 1H, CH arom, J = 8.8 Hz), 6.63 (d, 1H, CH arom, J = 5.2 Hz), 6.63 (s, 1H, CH arom), 6.03 (dd, 1H, CH-OPiv, J = 11.4, 3.0 Hz), 4.81 (dt, 1H, N-C H_2 , J = 13.6, 4.2 Hz), 2.88 (td, 1H, N- CH_2 , J = 12.1, 2.0 Hz), 2.56 (s, 3H, Ar– CH_3), 2.16– 2.25 (m, 2H, N-CH₂-CH₂), 1.75-1.9 (m, 2H, CH-CH₂), 1.36 (s, 9H, CH₃); 13 C NMR (CDCl₃, 100 MHz) δ 177.2 (O-CO), 168.04 (N-CO), 162.04 (d, 1C, C-F, ${}^{1}J_{C-F} = 248 \text{ Hz}$, 147.7 ($C-NO_2$), 142.0 ($C-NO_2$) N), 137.6 (*C*–CO), 134.8 (*C*–CH), 129.4 (d, 1C, *C*H arom, $^2J_{\text{C-F}}$ = 10.5 Hz), 127.5 (*C*H arom), 125.2 (d, 1C, CH arom, ${}^{2}J_{C-F} = 4.2 \text{ Hz}$, 120.9 (CH arom), 120.2 (CH arom), 114.8 (C-CH₃), 111.6 (d, 1C, CH arom, ${}^{3}J_{C-F} = 28.0 \text{ Hz}$), 72.0 (CH-OPiv), 46.6 (N- CH_2), 39.1 (C-(CH_3)₃), 32.1 (N- CH_2 - CH_2), 27.3 (3C, $(CH_3)_3$, 25.5 $(CH-CH_2)$, 19.9 $(Ar-CH_3)$; MS $(CI+NH_3)$ m/z: 446 (MH⁺+NH₃), 429 (MH⁺), 326 (MH⁺-PivOH); IR (cm⁻¹, CCl₄): 1735 (O–C=O), 1659 (N–C=O), 1529 (NO_2) , 1346 (NO_2) , 1139 (O-C=O).

4.1.18. 2,2-Dimethyl-propionic acid 1-(4-amino-2-methyl-benzoyl)-7-chloro-2,3,4,5-tetrahydro-1*H***-benzo|***b***|azepin-5-yl ester (15a).** To a solution of **14a** (0.15 g, 0.34 mmol) and HCl (0.2 mL) in ethanol (6 mL) at reflux was added SnCl₂ (0.32 g, 1.68 mmol). The mixture was refluxed for 2 h, cooled at room temperature, basified with saturated aqueous Na₂CO₃, extracted with ethyl acetate, dried over Na₂SO₄ and concentrated under reduced pressure. The residue was purified by flash column chromatography (silica gel, petroleum ether–AcOEt, 4:1) and recrystallised with petroleum ether to give compound **15a** (82% yield) as a white solid (mp 185–186 °C): ¹H

NMR (CDCl₃, 400 MHz) δ 7.22 (s, 1H, CH arom), 6.93 (d, 1H, CH arom, J = 8.0 Hz), 6.73 (d, 1H, CH arom, J = 8.0 Hz), 6.56 (d, 1H, CH arom, J = 8.0 Hz), 6.4– 6.46 (m, 2H, CH arom), 6.22 (d, 1H, CH arom, J = 8.0 Hz), 6.02 (d, 1H, CH–OPiv, J = 7.4 Hz), 4.84 (d, 1H, N–C H_2 , J = 8.4 Hz), 3.7 (br, 2H, NH), 3.78 (t, 1H, N-C H_2 , J = 12.0 Hz), 2.37 (s, 3H, Ar-C H_3), 2.05– 2.21 (m, 2H, N-CH₂-CH₂), 1.68-1.77 (m, 2H, CH- CH_2), 1.35 (s, 9H, CH_3); ¹³C NMR (CDCl₃, 100 MHz) δ 177.1 (O–CO), 170.4 (N–CO), 147.1 (C–NCO), 139.3 (C-NH₂), 137.7 (C-CO), 132.6 (C-Cl), 129.6 (CH arom), 128.7 (CH arom), 127.7 (CH arom), 125.6 (C-CH), 124.2 (CH arom), 116.6 (CH arom), 114.3 (C-CH₃), 111.9 (CH arom), 72.1 (CH-OPiv), 46.2 (N-CH₂), 39.0 (C-(CH₃)₃), 32.0 (N-CH₂-CH₂), 27.3 (3C, (CH₃)₃), 25.6 (CH–CH₂), 20.1 (Ar–CH₃); MS (CI+NH₃) m/z: 433 (MH⁺+NH₃), 432 (MH⁺+NH₃), 416 (MH⁺), 414 (MH⁺), 315 (MH⁺-PivOH), 313 (MH⁺-PivOH); IR (cm^{-1}, CCl_4) : 3400 (NH_2) , 1734 (O-C=O), 1651 (N-C=0), 1142 (O-C=0).

4.1.19. 2,2-Dimethyl-propionic acid 1-(4-amino-2-methylbenzovl)-7-fluoro-2,3,4,5-tetrahydro-1*H*-benzo[*b*|azepin-5-vl ester (15b). Using the same procedure as 15a, compound **15b** (78%) was obtained from **14b** (0.05 g, 0.13 mmol) as a colourless oil: 1 H NMR (CDCl₃, 400 MHz) δ 6.95 (d, 1H, CH arom, J = 9.6 Hz), 6.73 (d, 1H, CH arom, J = 8.0 Hz), 6.56–6.66 (m, 2H, CH arom), 6.39 (s, 1H, CH arom), 6.20 (d, 1H, CH arom, J = 8.4 Hz), 6.03 (d, 1H, CH-OPiv, J = 10.4 Hz), 4.84 (d, 1H, N-C H_2 , J = 13.6 Hz), 3.71 (br, 2H, N H_2), 2.77 (t, 1H, N-C H_2 , J = 12.2 Hz), 2.36 (s, 3H, Ar–C H_3), 2.13–2.15 (m, 2H, N–CH₂–C H_2), 1.69–1.84 (m, 2H, CH–C H_2), 1.34 (s, 9H, C H_3); ¹³C NMR (CDCl₃, 100 MHz) δ 177.1 (O– CO), 170.5 (N-CO), 161.3 (d, 1C, C-F, ${}^{1}J_{C-F}$ = 247 Hz), 150.3 (C-NCO), 147.0 (CH arom), 140.1 (C-NH₂), 137.6 (C-CO), 147.0 (CH arom), 140.1 (C-NH₂), 137.6 (C-CO), 129.9 (C-CH), 128.5 (CH arom), 116.5 (d, 1C, CH arom, ${}^{2}J_{C-F} = 12.6 \text{ Hz}$), 114.4 (d, 1C, CH arom, ${}^{2}J_{C-F} = 6.0 \text{ Hz}$), 112.2 (C-CH), 111.1 (C-CH₃), 110.7 (d, 1C, CH arom, ${}^{3}J_{C-F} = 26.4 \text{ Hz}$), 72.2 (CH OPin), 46.2 (N CH), 20.0 (CH) 26 Hz), 72.2 (CH-OPiv), 46.2 (N-CH₂), 39.0 (C- $(CH_3)_3$, 32.2 $(N-CH_2-CH_2)$, 27.3 $(3C, (CH_3)_3)$, 25.7 $(CH-CH_2)$, 19.3 $(Ar-CH_3)$; MS $(CI+NH_3)$ m/z: 399 (MH^+) , 298 $(MH^+-PivOH)$; IR (cm^{-1}, CCl_4) : 3399 (NH_2) , 1734 (O-C=O), 1649 (N-C=O), 1148 (O-C=O).

4.1.20. Tolvaptan (2). A solution of 2-methylbenzoyl chloride (0.05 g, 0.30 mmol) in CH₂Cl₂ (1 mL) was added dropwise to an ice-cooled solution of benzazepine 15a (0.05 g, 0.12 mmol) and triethylamine (0.07 mL, 0.05 g,0.48 mmol) in CH₂Cl₂ (1 mL). The mixture was stirred at room temperature for 1 h, neutralised with saturated aqueous Na₂CO₃, extracted with ethyl acetate, dried over Na₂SO₄ and concentrated under reduced pressure. The crude mixture was then dissolved in ethanol (1.5 mL) and 2 mL of a 2 N NaOH aqueous solution was added to the solution. The resulting solution was then heated at 50 °C for 2 h and cooled to room temperature. Water was then added to the reaction mixture and the insoluble material was removed by filtration. The solid thus obtained was washed with cold water and recrystallised from methanol/diethyl ether to furnish compound 1 (85% over 2 steps) as a white solid: mp 220–221 °C (lit.

225.9 °C) whose spectroscopic and analytical data were identical to those reported in the literature.

4.1.21. Fluoro-Tolvaptan (16). Using the same procedure as 2, compound 16 (quantitative yield) was obtained from **15b** (0.04 g, 0.10 mmol) as a white solid (mp 196–198 °C): ¹H NMR (CDCl₃, 400 MHz) δ 10.4 (br, 1H, NH), 7.70 (s,1H, CH arom), 7.4-7.63 (m, 6H, CH arom), 6.91-7.0 (m, 3H, CH arom), 5.92 (br, 1H, OH), 5.88 (d, 1H, CH-OH, J = 4.0 Hz), 5.06 (d, 1H, N- CH_2 , J = 10.0 Hz), 4.81 (d, 1H, N– $CH_{2'}$, J = 13.2 Hz), 2.83 (t, 1H, N-CH₂-C H_2 , J = 12.0 Hz), 2.53–2.56 (m, 1H, N-CH₂-CH₂), 2.50 (s, 6H, Ar-CH₃), 2.25-2.26 (m, 1H, CH–C H_2), 1.90–1.92 (m, 1H, CH–C H_2); ¹³C NMR (CDCl₃, 100 MHz) δ 167.8 (N–CO), 167.3 (N–CO), 160.8 (d, 1C, C–F, ${}^{1}J_{\text{C-F}}$ = 243 Hz), 145.6 (C-NCO), 139.2 (C-NCO), 136.9 (C-C(O)N), 136.0 (C-C(O)N), 135.8 (C-CH), 135.2 $(C-CH_3)$, 131.5 (C-CH₃), 130.4 (CH arom), 129.5 (d, 1C, CH arom, ${}^{3}J_{C-F}$ = 13.0 Hz), 128.2 (*CH* arom), 127.1 (*CH* arom), 126.3 (CH arom), 125.6 (CH arom), 120.8 (CH arom), 115.8 (CH arom), 113.3 (d, 1C, CH arom, ${}^{2}J_{C-F} =$ 23.1 Hz), 111.7 (d, 1C, CH arom, $^2J_{C-F} = 26.1$ Hz), 69.5 (CH-OH), 45.8 (N- CH_2), 35.4 (N- CH_2 - CH_2), 25.8 (CH–CH₂), 19.6 (Ar–CH₃), 19.2 (Ar–CH₃); MS (CI+NH₃) *m/z*: 433 (MH⁺); IR (cm⁻¹, CCl₄): 3490 (OH), 1640 (N-C=O), 1623 (N-C=O).

Acknowledgments

We thank CONACYT (Mexico) and CNRS (France) for its generous financial support to A.C.-V.

References and notes

- Schultz, C.; Link, A.; Leost, M.; Zaharevitz, D. W.; Gussio, R.; Sausville, E. A.; Meijer, L.; Kunick, C. J. Med. Chem. 1999, 42, 2909.
- (a) Watthey, J. W. H.; Gavin, T.; Desai, M. J. Med. Chem. 1984, 27, 816; (b) Watthey, J. W.; Stanton, J. L.; Desai, M.; Babiarz, J. E.; Finn, B. J. Med. Chem. 1985, 28, 1511.
- (a) Hori, M.; Fujimura, H.; Masuda, T.; Sawa, Y. Yakugaku Zasshi 1975, 95, 131; (b) Sawa, Y.; Kato, T.; Morimoto, T.; Hori, M.; Fujimura, H. Yakugaku Zasshi 1975, 95, 261; (c) Sawa, Y.; Kato, T.; Masuda, T.; Hori, M.; Fujimura, H. Chem. Pharm. Bull. 1975, 23, 1917.
- (a) Albright, J. D.; Reich, M. F.; Delos Santos, E. G.; Dusza, J. P.; Sum, F.-K.; Venkatesan, A. M.; Coupet, J.; Chan, P. S.; Ru, X.; Mazandarani, H.; Bailey, T. J. Med. Chem. 1998, 41, 2442; (b) Kondo, K.; Ogawa, H.; Shinohara, T.; Kimura, M.; Tanada, Y.; Kan, K.; Yamashita, H.; Nakamura, S.; Hirano, T.; Yamamura, Y.; Mori, T.; Tominaga, M.; Itai, A. J. Med. Chem. 2000, 43, 4388; (c) Matsubara, J.; Kitano, K.; Otsubo, K.; Kawano, Y.; Ohtani, T.; Bando, M.; Kido, M.; Uchida, M.; Tabusa, F. Tetrahedron 2000, 56, 4667; (d) Kondo, K.; Kan, K.; Tanada, Y.; Bando, M.; Shinohara, T.; Kurimura, M.; Ogawa, H.; Nakamura, S.; Hirano, T.; Yamamura, Y.; Kido, M.; Mori, T.; Tominaga, M. J. Med. Chem. 2002, 45, 3805.
- (a) Kondo, K.; Ogawa, H.; Yamashita, H.; Miyamoto, H.; Tanaka, M.; Nayaka, K.; Kitano, K.; Yamamura, Y.; Nakamura, S.; Onogawa, T.; Mori, T.; Tominaga, M.

- Bioorg. Med. Chem. 1999, 7, 1743; (b) Matsubara, J.; Morita, S.; Yamashita, H.; Otsubo, K.; Kitano, K.; Ohtani, T.; Kawano, Y.; Bando, M.; Kido, M.; Uchida, M.; Tabusa, F. Heterocycles 2001, 54, 131; (c) Yamashita, H.; Ohtani, T.; Morita, S.; Otsubo, K.; Kan, K.; Matsubara, J.; Kitano, K.; Kawano, Y.; Uchida, M.; Tabusa, F. Heterocycles 2002, 56, 123; (d) Sorbera, L. A.; Castañer, J.; Bayés, M.; Silvestre, J. Drugs Fut. 2002, 27, 350.
- Ogawa, H.; Yamashita, H.; Kondo, K.; Yamamura, Y.; Miyamoto, H.; Kan, K.; Kitano, K.; Tanaka, M.; Nayaka, K.; Nakamura, S.; Mori, T.; Tominaga, M.; Yabuuchi, Y. J. Med. Chem. 1996, 39, 3547.
- (a) Meyers, A. I.; Hutchings, R. H. Tetrahedron 1993, 34, 1807; (b) Nicholls, I. A.; Alewood, P. F. Bioorg. Chem. 1994, 22, 300; (c) Lowe, J. A.; Hageman, D. L.; Drozda, S. E.; McLean, S.; Bryce, D. K.; Crawford, R. T.; Zorn, S.; Morrone, J.; Bordner, J. J. Med. Chem. 1994, 37, 3789; For a review on the Beckmann reaction, see: (d) Gawley, R. E. Org. React. 1988, 35, 1.
- Liard, A.; Quiclet-Sire, B.; Saicic, R.; Zard, S. Z. Tetrahedron Lett. 1997, 38, 1759.

- Cordero-Vargas, A.; Pérez-Martin, I.; Quiclet-Sire, B.; Zard, S. Z. Org. Biomol. Chem. 2004, 2, 3018.
- Cordero-Vargas, A.; Quiclet-Sire, B.; Zard, S. Z. Tetrahedron Lett. 2004, 45, 7335.
- Cordero-Vargas, A.; Quiclet-Sire, B.; Zard, S. Z. Org. Lett. 2003, 20, 3717.
- (a) Zard, S. Z. Angew. Chem. Int. Ed. Engl. 1997, 36, 672;
 (b) Zard, S. Z. In Radicals in Organic Synthesis; Renaud, P., Sibi, M., Eds.; Wiley, VCH: Weinheim, 2001; pp 90–108.
- 13. Gagosz, F.; Zard, S. Z. Org. Lett. 2002, 4, 4345.
- (a) Francis, W. C. et al. J. Am. Chem. Soc. 1958, 80, 6238;
 (b) Sugg, E. E. et al. J. Org. Chem. 1985, 50, 5032.
- 15. Brown, H. C.; Heim, P. J. Org. Chem. 1973, 38, 912.
- Bhat, A.; Chang, H.-M.; Wallace, L. J.; Weinstein, D. M.; Shams, G.; Garris, C. C.; Hill, R. A. *Bioorg. Med. Chem.* 1998, 6, 271.
- (a) Hattori, K.; Matsumura, Y.; Miyazaki, T.; Maruoka, K.; Yamamoto, H. J. Am. Chem. Soc. 1981, 103, 7368; (b) Sasatani, S.; Miyazaki, T.; Maruoka, K.; Yamamoto, H. Tetrahedron Lett. 1983, 24, 4711.